

# Inflammatory signal transduction in Crohn's disease and novel therapeutic approaches

Catherine van Montfrans, Maikel Peppelenbosch<sup>\*</sup>, Anje A. te Velde,  
Sander van Deventer

*Laboratory for Experimental Internal Medicine, G2-133, Academic Medical Center,  
Meibergdreef 9, 1105 AZ Amsterdam, The Netherlands*

Received 4 March 2002; accepted 15 April 2002

---

## Abstract

Although signal transduction phenomena are well understood *in vitro*, their relevance for physiology and pathophysiology remains obscure. Recent years, however, have seen a plethora of attempts to link cell biochemistry and disease. At the Luxembourg conference it was attempted to link basal science to disease. In my presentation I focused on our research on stress activated kinases and other signalling elements in this respect in a variety of pro-inflammatory conditions. Knowledge of these pathways has now led to new therapeutic approaches, especially in Crohn's disease, which will be the subject of this commentary.

© 2002 Elsevier Science Inc. All rights reserved.

**Keywords:** Crohn's disease; MAP kinases; CD4<sup>+</sup> T cells; Therapy

---

## 1. Crohn's disease

Crohn's disease is an inflammatory chronic disorder of the digestive tract that is usually localised in the terminal ileum and the colon [1]. Clinical features consist of abdominal pain, diarrhoea and weight loss, and the disease typically starts in early adult life [2]. The incidence of Crohn's disease has increased in the past decades, and the current prevalence in the northern part of the world is 5–8‰ [2]. Crohn's disease and ulcerative colitis are considered as the inflammatory bowel diseases. Ulcerative colitis differs from Crohn's disease with regard to genetic background, immunological and clinical features, and this condition is not further addressed in this commentary. We will here discuss briefly the present views on etiology and treatment of Crohn's disease. The current notions on the most important cellular and molecular interactions that are involved in physiological immune responses will be considered in more detail (Fig. 1).

## 2. Etiology of Crohn's disease

Accumulating evidence suggests that Crohn's disease represents the outcome of three interactive factors: host susceptibility, enteric microflora and the mucosal immune system [3]. Host susceptibility is determined by environmental and genetic factors. Potentially relevant environmental factors include smoking and diet [4]. Genetic factors associated with IBD have been extensively studied and the first susceptibility gene for Crohn's disease has recently been identified. Mutations in the NOD2 gene, which encodes a protein with homology to plant disease resistance gene products, were found in 40–50% of Crohn's disease patients [5–7]. Homozygosity for a functionally relevant mutation within this NOD2 gene confers an extremely high risk for the development of Crohn's disease, and is considered to be the causative genetic defect in about 15% of the patients. It is tempting to speculate that NOD2 is involved in the defense against certain luminal bacteria: it is expressed by antigen presenting cells, such as monocytes and dendritic cells<sup>1</sup> and, following stimulation by bacterial LPS, activates nuclear factor- $\kappa$ B, a family of signal transduction proteins that regulates expression of

---

<sup>\*</sup> Corresponding author. Tel.: +31-20-566-8750; fax: +31-20-697-7192.

E-mail address: m.p.peppelenbosch@amc.uva.nl (M. Peppelenbosch).

Abbreviations: IBD, inflammatory bowel disease; LPS, lipopolysaccharides; Th, T helper; IL, interleukin; TGF- $\beta$ , transforming growth factor  $\beta$ ; IFN- $\gamma$ , interferon- $\gamma$ ; TNF- $\alpha$ , tumour necrosis factor.

<sup>1</sup> Braat H, unpublished results.

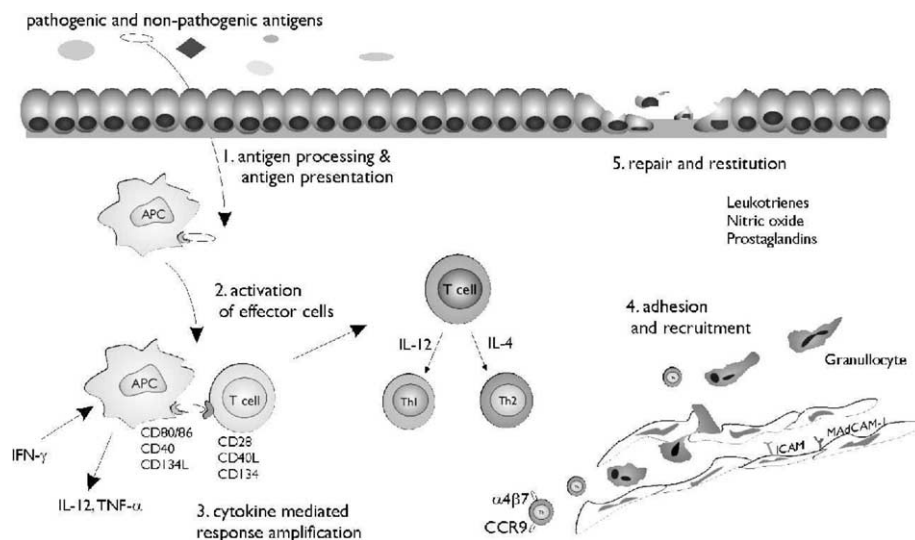


Fig. 1. Immune response in Crohn's disease and therapeutic targets for biological agents. Under inflammatory conditions, migrating dendritic cells transport antigens to regional lymph nodes. Presentation of antigens to naïve T cells in the presence of IL-12 results in the generation of Th1 effector cells that migrate into the intestine to mediate an inflammatory response. Targets for biological agents may be divided to according to the different phases of an immune response: (1) antigen and antigen presentation, (2) activation of effector T cells, (3) cytokine-mediated response amplification, (4) adhesion and recruitment, (5) repair and restitution.

many genes (i.e.  $\text{TNF-}\alpha$  and  $\text{IL-1}\beta$ ) involved in immune and inflammatory responses [5,8]. It is possible that early recognition of (pathogenic) bacteria is defective in Crohn's disease patients with mutant NOD2, leading to an abnormal T cell-mediated response, tissue inflammation and aberrant cytokine production. However, the precise functional consequences of the NOD2 mutations that are involved in the pathogenesis of Crohn's disease are not clear and need to be further characterised.

Many experimental and clinical studies have demonstrated that the enteric microflora is an essential cofactor in driving mucosal inflammation: germ-free mice do not develop experimental colitis [9,10], and diversion of the fecal stream following formation of a loop ileostomy has a beneficial effect in patients with Crohn's disease [11]. More direct evidence comes from studies by Duchmann *et al.*, who showed that mucosal, but not peripheral blood,  $\text{CD4}^+$  T cells from IBD patients proliferate when exposed to autologous intestinal bacteria [12,13].

The mucosal immune system is activated upon exposure to a pathogenic microorganism. Initially, innate immune cells, such as macrophages, dendritic cells and natural killer cells are recruited to eliminate pathogens by uptake and destruction or degradation [14]. Macrophages and dendritic cells are activated and can present the products of pathogen degradation to antigen reactive T cells resulting in an adaptive immune response. T cells, in particular  $\text{CD4}^+$  T cells, play multiple roles as effector and regulatory cells in intestinal inflammation: a subset of naïve  $\text{CD45RB}^{\text{high}} \text{CD4}^+$  cells [15] has been shown to induce experimental colitis after transfer to severe combined immunodeficient (SCID) mice [16]. In addition, the number of activated mucosal T cells secreting pro-inflammatory

cytokines that mediate interactions between cells, such as  $\text{IFN-}\gamma$ , is increased in active Crohn's disease [17,18]. Finally,  $\text{CD4}^+$  cell depletion ameliorates inflammation in experimental colitis [19] and in Crohn's disease [20]. By comparison, whereas  $\text{CD8}^+$  cells are important in first line defence in the epithelium [21], they do not appear to play a decisive pathologic role in intestinal inflammation [22,23].

Based on these observations, the current working model of the pathophysiology of Crohn's disease is that of an uncontrolled abnormal immune response of  $\text{CD4}^+$  cells to enteric bacteria in genetically susceptible individuals.

### 3. Therapy of Crohn's disease

Currently there is no curative treatment for patients with Crohn's disease. For several decades corticosteroids were the only potent anti-inflammatory agents available for treatment of active Crohn's disease [24]. Corticosteroids act rapidly and have a wide range of anti-inflammatory activities, but do not alter the natural course of the disease, do not consistently heal intestinal ulcers, and are associated with many side effects [25]. The use of immunomodulatory therapy, as azathioprine and methotrexate has provided alternative therapeutic options [26–28]. However, the administration of these agents is limited by inconstant efficacy, a relatively slow onset of action, inadequate selectivity and substantial short- and long-term toxicity.

New biological therapeutic strategies are based on the increased knowledge of the pathophysiology of Crohn's disease. Biological agents include recombinant cytokines, cytokine receptor antagonists, chimeric and humanised antibodies, antisense oligonucleotides and viral as well

as non-viral vectors for gene transfer. Presently, of this growing range of biologic therapies antagonism of TNF- $\alpha$  with the monoclonal TNF- $\alpha$  antibody infliximab is the only approach that has been approved for clinical use. Infliximab is effective and well tolerated in patients with moderate to severe Crohn's disease that is resistant to conventional treatment [29] and can maintain remission up to 44 weeks if administered by repeated intravenous infusions [30]. The rapid healing of enterocutaneous fistulas is a good example of the anti-inflammatory activity of infliximab in Crohn's disease [31]. Traditional therapeutic agents have limited mucosal healing effects, and in the older studies endoscopic severity of ileocolonic Crohn's disease did not correlate well with therapeutic effects as measured by clinical parameters. However, the clinical improvement following infliximab therapy has been consistently associated with mucosal healing [32,33]. The long-term efficacy and safety as well as the precise mode of action of infliximab are currently under investigation. However, because of the immunogenicity of monoclonal antibodies and risk of side effects such as opportunistic infections and malignancy, infliximab may be best suited as a bridge to more long-term maintenance strategies. Furthermore, gene therapy has emerged as a new and promising method for maintenance of remissions in chronic inflammatory diseases, but its clinical application is still far away.

#### 4. Intestinal immune response in physiological conditions: peripheral tolerance

Although the intestinal epithelium forms a physical barrier for pathogens, bacterial antigens are continuously sampled by gut mucosal dendritic cells, the major intestinal antigen presenting cells [34], and presented to T cells [35]. The mucosal immune system needs to discriminate pathogens from dietary antigens and commensal bacteria present in the gut lumen. Indeed, a large pool of immune cells is present within the mucosal epithelium and lamina propria. The continuous presence of potential pathogens in close proximity to immune cells must be tightly controlled to ensure that pathologic inflammation does not develop. Control is provided by mechanisms of tolerance. The concept of tolerance includes, by definition, any mechanism by which a potentially injurious immune response is prevented, suppressed, or shifted to a non-injurious class of immune response [36]. Central tolerance refers to thymic deletion of self-reactive cells, which is not complete and self-reactive cells escape to the periphery. Several mechanisms are involved in maintaining peripheral tolerance, i.e. *anergy* (functional inactivity), *apoptosis* (programmed cell death) and the involvement of *tolerogenic dendritic cells* or *regulatory T cells* [37]. These mechanisms can interfere during all phases of the immune response and are probably all necessary to control (1) self-

reactive cells, (2) cells reactive to non-pathogenic microorganisms and (3) cells that become activated following a normal immune response to a foreign antigen.

#### 5. Physiological immune response to pathogens

The intestinal mucosa contains a large number of dendritic cells for uptake, processing and transport of antigens to the regional lymph nodes. Dendritic cells critically steer immune responses through interactions with T cells, requiring close proximity and crosstalk of these two cell types. The decision between immunity and tolerance is taken by dendritic cells: they induce the development of naïve T cells into effector or regulatory T cells, or can induce anergy/apoptosis of T cells [38–40].

Dendritic cells express pattern recognition receptors (e.g. Toll-like receptors) [41] that recognise pathogen-associated molecules (e.g. LPS) from invading microbes. Such activation of pattern recognition receptors generally increases the immune-stimulatory capacities of dendritic cells. After antigen uptake, activated and maturing dendritic cells start to express the chemokine receptor CCR7. Dendritic cells migrate to the lymph nodes, where specialised high endothelial venules express the CCR7 ligand CCL2 [42]. CCR7 is also present on naïve T cells [43,44] and directs, together with the lymph node homing receptor L-selectin [45] the migration of naïve T cells to the regional lymph nodes. The passage across the high endothelial venules is a multistep process that involves selectin-supported rolling, followed by a triggering event, and firm integrin-mediated adhesion [46,47]. Chemokines are a family of leukocyte chemoattractants that regulate both inflammatory cell recruitment and homeostatic trafficking of leukocytes.

When dendritic cells present antigens to naïve T cells in a lymph node, the T cells differentiate into effector/memory CD4<sup>+</sup> or cytotoxic CD8<sup>+</sup> cells. Based on their cytokine secretion profile, CD4<sup>+</sup> T cells differentiate into at least two subsets of helper cells, Th1 and Th2 cells [48]. Th1 cells produce the pro-inflammatory cytokines IL-2 and IFN- $\gamma$  and protect against intracellular pathogens (cellular response), whereas Th2 cells selectively secrete IL-4, IL-5 and IL-13, and counter extracellular pathogens through production of antibody by B cells (humoral response). Although both subsets of T helper cells can also produce the two pivotal immunoregulatory cytokines, IL-10 and TGF- $\beta$ , regulatory T cells, which are discussed more fully below, are the main producers of these cytokines. The cytokine microenvironment to which naïve T cells are exposed is the primary determining factor for the differentiation towards a Th1 or Th2 cell type [49]: IL-12 with support of IL-18, both produced by antigen presenting cells, induce Th1 differentiation [50,51] and IL-4, produced by NK cells, mast cells, basophils and mature CD4<sup>+</sup> cells [52], drives Th2 differentiation [53]. Others factors

such as antigen density [54], major histocompatibility complex haplotypes [55], type of antigen presenting cell [56], and costimulatory factors [57] also influence the selective development of Th1/Th2 cells and regulatory T cells.

After local priming of T cells in the presence of—high levels of—IL-12 and IL-18, the differentiated effector Th1 cells and memory T cells recirculate preferentially to the intestinal mucosa. The integrin  $\alpha 4\beta 7$ , expressed on these T cells, is the principal gut homing receptor and functions at several steps in the adhesion cascade by interacting with the mucosal addressin MAdCAM-1 present on endothelial cells [58,59]. In addition, the chemokine TECK and its receptor CCR9 are implicated in the migration of  $\alpha 4\beta 7^+$  memory T cells to the small intestine [60]. Once arrived in the intestine the Th1 effector cells mount an inflammatory response on encounter of the antigen, and this response can be amplified by freshly recruited immature dendritic cells. After elimination of the pathogen, the bulk of the effector T cells become redundant and most of them disappear, in large part through apoptosis. However, a small proportion survives to become long-lived memory cells [61]. Thus, whereas induction of anergy and apoptosis serve to reduce the number of T cells that can respond to mucosal antigens, this mechanism is not sufficient for elimination of all potentially reactive T cells. Cells that escape these mechanisms of peripheral tolerance need to be controlled by tolerogenic dendritic cells and/or regulatory T cells.

## 6. What is wrong with the mucosal immune system in Crohn's disease?

Clinical and experimental studies have provided evidence for several defects of the mucosal immune response in Crohn's disease. Based on the predominance of IL-2 and IFN- $\gamma$  secreting mucosal T cells and the specific expression of Th1 driving cytokines IL-12 and IL-18 in the intestinal mucosa, Crohn's disease is considered to be a prototype Th1 disease [18,62–64]. Apparently, mechanisms of peripheral tolerance are not effective in down-regulating this Th1 cell-mediated inflammation, and two studies have reported that mucosal T cells in patients with Crohn's disease are resistant to apoptosis [65,66]. Our laboratory has recently demonstrated that the number of dendritic cells producing IL-12 and IL-18 in the intestinal mucosa is increased in patients with Crohn's disease, indicating the enhanced activation of the immune system in the intestine.<sup>2</sup>

In the intestinal mucosa of healthy individuals, CD4<sup>+</sup> cells have been shown to contribute to down-regulation of effector T cells, via production of IL-10 and TGF- $\beta$  [67]. Conflicting reports exist concerning suppressor/regulatory cell activity by lamina propria cells [68,69] and peripheral blood mononuclear cells [70,71] in Crohn's

disease patients. Crohn's disease is characterised by a relatively reduced production of bioactive IL-10 in the lamina propria, and the resulting IL-10 concentrations seem insufficient to down-regulate pro-inflammatory cytokines [72,73]. A low ileal IL-10 concentration is associated with early endoscopic recurrence of disease after surgery in patients with Crohn's disease [74]. Our laboratory has recently found that mucosal memory (CD45RB<sup>low</sup> CD45RO<sup>+</sup>) CD4<sup>+</sup> cells from patients with Crohn's disease are less abundant than in controls and produce less IL-10 [75]. Conversely, increased TGF- $\beta$ 1 expression was found in affected mucosa of patients with active Crohn's disease [76,77], and it was shown that mucosal T cells from IBD patients were insensitive to the regulatory effects of TGF- $\beta$  because they overexpress SMAD7, an inhibitor of TGF- $\beta$  signalling [78].

These data suggest that in Crohn's disease several defects in the mechanisms of peripheral tolerance, including resistance to T cell apoptosis and disrupted regulatory T cell function can exist leading to a final common pathway of Th1-mediated intestinal inflammation. It is logical to assume that interference with underlying signalling pathways will have clinical benefit in Crohn's disease.

## 7. MAP kinases

MAP kinases are a family of evolutionary well-conserved proteins that are expressed in all cells, and regulate survival, growth and inflammation. MAP kinases are activated by phosphorylation of both a threonine and serine residue and subsequently phosphorylate downstream kinases and transcription factors. This leads to altered transcription of genes that regulate growth, apoptosis or inflammation, or altered post-transcriptional events like translation or mRNA degradation. In mammalian cells, three main MAP kinase pathways are of importance, i.e. p38 MAP kinase, p42/p44 MAP kinase and the c-Jun-N-terminal kinase. Of these, the p38 MAP kinase pathway has attracted most attention as a therapeutic target, because it was shown to mediate cellular responses to stress, induced by signals such as cytokines and bacterial components. Moreover, the p38 MAP kinase pathway is activated in LPS-infused human volunteers [79] and blocking p38 MAP inhibitors in such volunteers strongly reduced clinical symptoms [80].

More recently, the other MAP kinases have been reported to mediate inflammation and we investigated whether inhibition of MAP kinases was beneficial in Crohn's disease. Inhibition of JNK and p38 MAPK activation with CNI-1493, a guanilylhydrazone, was tested in 12 patients with severe Crohn's disease. Colonic biopsies displayed enhanced JNK and p38 MAPK activation. CNI-1493 inhibition of both JNK and p38 phosphorylation was observed *in vitro*. Treatment resulted in diminished JNK phosphorylation and TNF production as well as

<sup>2</sup>te Velde A, unpublished results.



significant clinical benefit and rapid endoscopic ulcer healing. No serious adverse events were noted. A CDAI decrease of 120 at week 4 ( $P = 0.005$ ) and 146.5 at week 8 ( $P = 0.005$ ) was observed. A clinical response was seen in 67% of patients at 4 weeks and 58% at 8 weeks. Clinical remission was observed in 25% of patients at week 4 and 42% at week 8. Endoscopic improvement occurred in all but one patient. Response was seen in three of six infliximab failures, two of which showed remission. Fistulae healing occurred in four of five patients, and steroids were tapered in 89% of patients [81]. In conclusion, knowledge of signal transduction pathways has now led to the realisation that inflammatory MAPKs are critically involved in the pathogenesis of Crohn's disease and their inhibition provides a novel and powerful therapeutic option.

## References

- [1] Targan SR, Fergus S. In: Retford DC, editor. From bench to bedside. Baltimore: Williams & Wilkins, 1994. p. 795.
- [2] Lapidus A, Bernell O, Hellers G, Persson PG, Lofberg R. Incidence of Crohn's disease in Stockholm County 1955–1989. *Gut* 1997;41:480–6.
- [3] Fiocchi C. Inflammatory bowel disease: etiology and pathogenesis. *Gastroenterology* 1998;115:182–205.
- [4] Somerville KW, Logan RF, Edmond M, Langman MJ. Smoking and Crohn's disease. *Br Med J (Clin Res Ed)* 1984;289:954–6.
- [5] Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, Britton H, Moran T, Karaliuskas R, Duerr RH, Achkar JP, Brant SR, Bayless TM, Kirschner BS, Hanauer SB, Nunez G, Cho JH. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 2001;411:603–6.
- [6] Hugot JP, Chamaillard M, Zouali H, Lesage S, Cezard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, Binder V, Finkel Y, Cortot A, Modigliani R, Laurent-Puig P, Gower-Rousseau C, Macry J, Colombel JF, Sahbatou M, Thomas G. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 2001;411:599–603.
- [7] Hampe J, Cuthbert A, Croucher PJ, Mirza MM, Mascheretti S, Fisher S, Frenzel H, King K, Hasselmeier A, MacPherson AJ, Bridger S, van Deventer S, Forbes A, Nikolaus S, Lennard-Jones JE, Foelsch UR, Krawczak M, Lewis C, Schreiber S, Mathew CG. Association between insertion mutation in NOD2 gene and Crohn's disease in German and British populations. *Lancet* 2001;357:1925–8.
- [8] Ghosh S, May MJ, Kopp EB. NF $\kappa$ B and rel proteins: evolutionarily conserved mediators of immune response. *Annu Rev Immunol* 1998;16:225–60.
- [9] Kuhn R, Lohler J, Rennick D, Rajewsky K, Muller W. Interleukin-10-deficient mice develop chronic enterocolitis. *Cell* 1993;75:263–74.
- [10] Aranda R, Sydora BC, McAllister PL, Binder SW, Yang HY, Targan SR, Kronenberg M. Analysis of intestinal lymphocytes in mouse colitis mediated by transfer of CD4<sup>+</sup>, CD45RB<sup>high</sup> T cells to SCID recipients. *J Immunol* 1997;158:3464–673.
- [11] Rutgeerts P, Goboos K, Peeters M, Hiele M, Penninckx F, Aerts R, Kerremans R, Vantrappen G. Effect of faecal stream diversion on recurrence of Crohn's disease in the neoterminal ileum. *Lancet* 1991;338:771–4.
- [12] Duchmann R, Kaiser I, Hermann E, Mayet W, Ewe K, Meyer zum Buschenfelde KH. Tolerance exists towards resident intestinal flora but is broken in active inflammatory bowel disease (IBD). *Clin Exp Immunol* 1995;102:448–55.
- [13] Duchmann R, Marker-Hermann E, Meyer zum Buschenfelde KH. Bacteria-specific T-cell clones are selective in their reactivity towards different enterobacteria or *H. pylori* and increased in inflammatory bowel disease. *Scand J Immunol* 1996;44:71–9.
- [14] Aderem A, Underhill DM. Mechanisms of phagocytosis in macrophages. *Annu Rev Immunol* 1999;17:593–623.
- [15] Lee WT, Yin XM, Vitetta ES. Functional and ontogenetic analysis of murine CD45Rhi and CD45Rlo CD4<sup>+</sup> T cells. *J Immunol* 1990;144:3288–95.
- [16] Powrie F, Leach MW, Mauze S, Caddle LB, Coffman RL. Phenotypically distinct subsets of CD4<sup>+</sup> T cells induce or protect from chronic intestinal inflammation in C. B-17 SCID mice. *Int Immunol* 1993;5:1461–71.
- [17] Fais S. Lymphocyte traffic and adhesion molecules in the gut. *Ital J Gastroenterol* 1991;23:395.
- [18] Fuss IJ, Neurath M, Boirivant M, Klein JS, de la Motte C, Strong SA, Fiocchi C, Strober W. Disparate CD4<sup>+</sup> lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease: Crohn's disease LP cells manifest increased secretion of IFN- $\gamma$ , whereas ulcerative colitis LP cells manifest increased secretion of IL-5. *J Immunol* 1996;157:1261–70.
- [19] Okamoto S, Watanabe M, Yamazaki M, Yajima T, Hayashi T, Ishii H, Mukai M, Yamada T, Watanabe N, Jameson BA, Hibi T. A synthetic mimetic of CD4 is able to suppress disease in a rodent model of immune colitis. *Eur J Immunol* 1999;29:355–66.
- [20] Stronkhorst A, Radema S, Yong SL, Bijl H, ten Berge IJ, Tytgat GN, van Deventer SJ. CD4 antibody treatment in patients with active Crohn's disease: a phase I dose finding study. *Gut* 1997;40:320–7.
- [21] Hayday A, Theodoridis E, Ramsburg E, Shires J. Intraepithelial lymphocytes: exploring the Third Way in immunology. *Nat Immunol* 2001;2:997–1003.
- [22] Simpson SJ, Mizoguchi E, Allen D, Bhan AK, Terhorst C. Evidence that CD4<sup>+</sup>, but not CD8<sup>+</sup> T cells are responsible for murine interleukin-2-deficient colitis. *Eur J Immunol* 1995;25:2618–25.
- [23] Davidson NJ, Leach MW, Fort MM, Thompson-Snipes L, Kuhn R, Muller W, Berg DJ, Rennick DM. T helper cell 1-type CD4<sup>+</sup> T cells, but not B cells, mediate colitis in interleukin 10-deficient mice. *J Exp Med* 1996;184:241–51.
- [24] Summers RW, Switz DM, Sessions JTJ, Beckett JM, Best WR, Kern FJ, Singleton JW. National Cooperative Crohn's Disease Study: results of drug treatment. *Gastroenterology* 1979;77:847–69.
- [25] Rutgeerts PJ. The limitations of corticosteroid therapy in Crohn's disease. *Aliment Pharmacol Ther* 2001;15:1515–25.
- [26] Sandborn WJ, McLeod R, Jewell DP. Medical therapy for induction and maintenance of remission in pouchitis: a systematic review. *Inflamm Bowel Dis* 1999;5:33–9.
- [27] Feagan BG, Rochon J, Fedorak RN, Irvine EJ, Wild G, Sutherland L, Steinhart AH, Greenberg GR, Gillies R, Hopkins M, et al. Methotrexate for the treatment of Crohn's disease: The North American Crohn's Study Group Investigators. *N Engl J Med* 1995;332:292–7.
- [28] Feagan BG, Fedorak RN, Irvine EJ, Wild G, Sutherland L, Steinhart AH, Greenberg GR, Koval J, Wong CJ, Hopkins M, Hanauer SB, McDonald JW. A comparison of methotrexate with placebo for the maintenance of remission in Crohn's disease: The North American Crohn's Study Group Investigators. *N Engl J Med* 2000;342:1627–32.
- [29] Targan SR, Hanauer SB, van Deventer SJH, Mayer L, Present DH, Braakman T, Dewoody KL, Schaible TF, Rutgeerts PJ. A short-term study of chimeric monoclonal antibody cA2 to tumour necrosis factor alpha for Crohn's disease. *N Engl J Med* 1997;337:1029–35.
- [30] Rutgeerts P, D'Haens G, Targan S, Vasilias E, Hanauer SB, Present DH, Mayer L, van Hogezaand RA, Braakman T, DeWoody KL, Schaible TF, van Deventer SJH. Efficacy and safety of retreatment with anti-tumour necrosis factor antibody (infliximab) to maintain remission in Crohn's disease. *Gastroenterology* 1999;117:761–9.
- [31] Present DH, Rutgeerts P, Targan SR, Hanauer SB, Mayer L, van Hogezaand RA, Podolsky DK, Sands BE, Braakman T, DeWoody KL, Schaible TF, van Deventer SJH. Infliximab for the treatment of fistulas in patients with Crohn's disease. *N Engl J Med* 1999;340:1398–405.

- [32] D'Haens G, van Deventer S, van Hogezaand R, Chalmers D, Kothe C, Baert F, Braakman T, Schaible T, Geboes K, Rutgeerts P. Endoscopic and histological healing with infliximab anti-tumour necrosis factor antibodies in Crohn's disease: a European multicenter trial. *Gastroenterology* 1999;116:1029–34.
- [33] Rutgeerts P. Infliximab is the drug we have been waiting for in Crohn's disease. *Inflamm Bowel Dis* 2000;6:132–6.
- [34] Harper HM, Cochrane L, Williams NA. The role of small intestinal antigen-presenting cells in the induction of T-cell reactivity to soluble protein antigens: association between aberrant presentation in the lamina propria and oral tolerance. *Immunology* 1996;89:449–56.
- [35] Rescigno M, Urbano M, Valzasina B, Francolini M, Rotta G, Bonasio R, Granucci F, Kraehenbuhl JP, Ricciardi-Castagnoli P. Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nat Immunol* 2001;2:361–7.
- [36] Weiner HL. Oral tolerance, an active immunologic process mediated by multiple mechanisms. *J Clin Invest* 2000;106:935–7.
- [37] Van Parijs L, Abbas AK. Homeostasis and self-tolerance in the immune system: turning lymphocytes off. *Science* 1998;280:243–8.
- [38] Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ, Pulendran B, Palucka K. Immunobiology of dendritic cells. *Annu Rev Immunol* 2000;18:767–811.
- [39] Huang FP, Platt N, Wykes M, Major JR, Powell TJ, Jenkins CD, MacPherson GG. A discrete subpopulation of dendritic cells transports apoptotic intestinal epithelial cells to T cell areas of mesenteric lymph nodes. *J Exp Med* 2000;191:435–43.
- [40] Jonuleit H, Schmitt E, Steinbrink K, Enk AH. Dendritic cells as a tool to induce anergic and regulatory T cells. *Trends Immunol* 2001;22:394–400.
- [41] Rock FL, Hardiman G, Timans JC, Kastelein RA, Bazan JF. A family of human receptors structurally related to *Drosophila* Toll. *Proc Natl Acad Sci USA* 1998;95:588–93.
- [42] Chan VW, Kothakota S, Rohan MC, Panganiban-Lustan L, Gardner JP, Wachowicz MS, Winter JA, Williams LT. Secondary lymphoid-tissue chemokine (SLC) is chemotactic for mature dendritic cells. *Blood* 1999;93:3610–6.
- [43] Campbell JJ, Bowman EP, Murphy K, Youngman KR, Siani MA, Thompson DA, Wu L, Zlotnik A, Butcher EC. 6-C-kine (SLC), a lymphocyte adhesion-triggering chemokine expressed by high endothelium, is an agonist for the MIP-3beta receptor CCR7. *J Cell Biol* 1998;141:1053–9.
- [44] Gunn MD, Tangemann K, Tam C, Cyster JG, Rosen SD, Williams LT. A chemokine expressed in lymphoid high endothelial venules promotes the adhesion and chemotaxis of naive T lymphocytes. *Proc Natl Acad Sci USA* 1998;95:258–63.
- [45] Picker LJ, Treer JR, Ferguson-Darnell B, Collins PA, Buck D, Terstappen LW. Control of lymphocyte recirculation in man. I. Differential regulation of the peripheral lymph node homing receptor L-selectin on T cells during the virgin to memory cell transition. *J Immunol* 1993;150:1105–21.
- [46] Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell* 1994;76:301–14.
- [47] Butcher EC, Picker LJ. Lymphocyte homing and homeostasis. *Science* 1996;272:60–6.
- [48] Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. *Annu Rev Immunol* 1989;7:145–73.
- [49] Abbas AK, Murphy KM, Sher A. Functional diversity of helper T lymphocytes. *Nature* 1996;383:787–93.
- [50] Okamura H, Tsutsi H, Komatsu T, Yutsudo M, Hakura A, Tanimoto T, Torigoe K, Okura T, Nukada Y, Hattori K, et al. Cloning of a new cytokine that induces IFN-gamma production by T cells. *Nature* 1995;378:88–91.
- [51] Kohno K, Kataoka J, Ohtsuki T, Suemoto Y, Okamoto I, Usui M, Ikeda M, Kurimoto M. IFN-gamma-inducing factor (IGIF) is a costimulatory factor on the activation of Th1 but not Th2 cells and exerts its effect independently of IL-12. *J Immunol* 1997;158:1541–50.
- [52] Liew FY. Th1 and Th2 cells: a historical perspective. *Nat Immunol Rev* 2002;2:55–60.
- [53] Swain SL, Weinberg AD, English M, Huston G. IL-4 directs the development of Th2-like helper effectors. *J Immunol* 1990;145:3796–806.
- [54] Hsieh CS, Heimberger AB, Gold JS, O'Garra A, Murphy KM. Differential regulation of T helper phenotype development by interleukins 4 and 10 in an alpha beta T-cell-receptor transgenic system. *Proc Natl Acad Sci USA* 1992;89:6065–9.
- [55] Constant S, Pfeiffer C, Woodard A, Pasqualini T, Bottomly K. Extent of T cell receptor ligation can determine the functional differentiation of naive CD4<sup>+</sup> T cells. *J Exp Med* 1995;182:1591–6.
- [56] Rissoan MC, Soumelis V, Kadowaki N, Grouard G, Briere F, de Waal Malefyt R, Liu YJ. Reciprocal control of T helper cell and dendritic cell differentiation. *Science* 1999;283:1183–6.
- [57] Howland KC, Ausubel LJ, London CA, Abbas AK. The roles of CD28 and CD40 ligand in T cell activation and tolerance. *J Immunol* 2000;164:4465–70.
- [58] Berlin C, Bargatz RF, Campbell JJ, von Andrian UH, Szabo MC, Hasslen SR, Nelson RD, Berg EL, Erlandsen SL, Butcher EC. Alpha 4 integrins mediate lymphocyte attachment and rolling under physiologic flow. *Cell* 1995;80:413–22.
- [59] Bargatz RF, Jutila MA, Butcher EC. Distinct roles of L-selectin and integrins alpha 4 beta 7 and LFA-1 in lymphocyte homing to Peyer's patch-HEV *in situ*: the multistep model confirmed and refined. *Immunity* 1995;3:99–108.
- [60] Zabel BA, Agace WW, Campbell JJ, Heath HM, Parent D, Roberts AI, Ebert EC, Kassam N, Qin S, Zovko M, LaRosa GJ, Yang LL, Soler D, Butcher EC, Ponath PD, Parker CM, Andrew DP. Human G protein-coupled receptor GPR-9-6/CC chemokine receptor 9 is selectively expressed on intestinal homing T lymphocytes, mucosal lymphocytes, and thymocytes and is required for thymus-expressed chemokine-mediated chemotaxis. *J Exp Med* 1999;190:1241–56.
- [61] Sprent J, Surh CD. T cell memory. *Annu Rev Immunol* 2002;20:551–79.
- [62] Parronchi P, Romagnani P, Annunziato F, Sampognaro S, Becchio A, Giannarini L, Maggi E, Pupilli C, Tonelli F, Romagnani S. Type 1 T-helper cell predominance and interleukin-12 expression in the gut of patients with Crohn's disease. *Am J Pathol* 1997;150:823–32.
- [63] Monteleone G, Biancone L, Marasco R, Morrone G, Marasco O, Luzzza F, Pallone F. Interleukin 12 is expressed and actively released by Crohn's disease intestinal lamina propria mononuclear cells. *Gastroenterology* 1997;112:1169–78.
- [64] Monteleone G, Trapasso F, Parrello T, Biancone L, Stella A, Iuliano R, Luzzza F, Fusco A, Pallone F. Bioactive IL-18 expression is up-regulated in Crohn's disease. *J Immunol* 1999;163:143–7.
- [65] Ina K, Itoh J, Fukushima K, Kusugami K, Yamaguchi T, Kyokane K, Imada A, Binion DG, Musso A, West GA, Dobrea GM, McCormick TS, Lapetina EG, Levine AD, Ottaway CA, Fiocchi C. Resistance of Crohn's disease T cells to multiple apoptotic signals is associated with a Bcl-2/Bax mucosal imbalance. *J Immunol* 1999;163:1081–90.
- [66] Boirivant M, Marini M, Di Felice G, Pronio AM, Montesani C, Tersigni R, Strober W. Lamina propria T cells in Crohn's disease and other gastrointestinal inflammation show defective CD2 pathway-induced apoptosis. *Gastroenterology* 1999;116:557–65.
- [67] Khoo UY, Proctor IE, Macpherson AJ. CD4<sup>+</sup> T cell down-regulation in human intestinal mucosa: evidence for intestinal tolerance to luminal bacterial antigens. *J Immunol* 1997;158:3626–34.
- [68] Fiocchi C, Youngman KR, Farmer RG. Immunoregulatory function of human intestinal mucosa lymphoid cells: evidence for enhanced suppressor cell activity in inflammatory bowel disease. *Gut* 1983;24:692–701.
- [69] Goodacre RL, Bienenstock J. Reduced suppressor cell activity in intestinal lymphocytes from patients with Crohn's disease. *Gastroenterology* 1982;82:653–8.

- [70] MacDermott RP, Bragdon MJ, Thurmond RD. Peripheral blood mononuclear cells from patients with inflammatory bowel disease exhibit normal function in the allogeneic and autologous mixed leukocyte reaction and cell-mediated lympholysis. *Gastroenterology* 1984;86:476–84.
- [71] Hodgson HJ, Wands JR, Isselbacher KJ. Decreased suppressor cell activity in inflammatory bowel disease. *Clin Exp Immunol* 1978;32:451–8.
- [72] Autschbach F, Braunstein J, Helmke B, Zuna I, Schürmann G, Niemi ZI, Wallich R, Otto HF, Meuer SC. *In situ* expression of interleukin 10 in noninflamed human gut and in inflammatory bowel disease. *Am J Pathol* 1998;153:121–30.
- [73] Gasche C, Bakos S, Dejaco C, Tillinger W, Zakeri S, Reinisch W. IL-10 secretion and sensitivity in normal human intestine and inflammatory bowel disease. *J Clin Immunol* 2000;20:362–70.
- [74] Meresse B, Rutgeerts P, Malchow H, Dubucquoi S, Dessaint JP, Cohard M, Colombel JF, Desreumaux P. Low ileal interleukin 10 concentrations are predictive of endoscopic recurrence in patients with Crohn's disease. *Gut* 2002;50:25–8.
- [75] ten Hove T, The FO, Bruggeman JP, Slors F, van Deventer SJH, te Velde AA. Marked difference of CD4<sup>+</sup>CD45RB<sup>high</sup> expression by gut derived T lymphocytes in Crohn's disease and ulcerative colitis (abstract). *Gastroenterology* 2001;120:A526.
- [76] Babyatsky MW, Rossiter G, Podolsky DK. Expression of transforming growth factors alpha and beta in colonic mucosa in inflammatory bowel disease. *Gastroenterology* 1996;110:975–84.
- [77] di Mola FF, Friess H, Scheuren A, Di Sebastiano P, Graber H, Egger B, Zimmermann A, Korc M, Buchler MW. Transforming growth factor-beta s and their signaling receptors are coexpressed in Crohn's disease. *Ann Surg* 1999;229:67–75.
- [78] Monteleone G, Kumberova A, Croft NM, McKenzie C, Steer HW, MacDonald TT. Blocking Smad7 restores TGF-beta1 signaling in chronic inflammatory bowel disease. *J Clin Invest* 2001;108:601–9.
- [79] van den Blink B, Branger J, Weijer S, van Deventer SJH, van der Poll T, Peppelenbosch MP. Human endotoxemia activates p38 MAP kinase and p42/p44 MAP kinase, but not c-Jun N-terminal kinase. *Mol Med* 2001;7:755–60.
- [80] Branger J, van den Blink B, Weijer S, Madwed J, Bos CL, Gupta A, Polmar AH, Olszyna DP, Hack CE, van Deventer SJH, Peppelenbosch MP, van der Poll T. Anti-inflammatory effects of a p38 mitogen activated protein kinase inhibitor, BIRB 796 BS, during human endotoxemia. *J Immunol* 2002;168:4070–7.
- [81] Hommes D, van den Blink B, Plasse T, Bartelsman J, Xu C, MacPherson B, Tytgat G, Peppelenbosch M, van Deventer S. Inhibition of stress-activated MAP kinases induces clinical improvement in moderate to severe Crohn's disease. *Gastroenterology* 2002;122:7–14.